



Washington State Toxics Monitoring and Resident Fish Movement in the Columbia River

William Hobbs

Environmental Assessment Program, Toxic Studies Unit

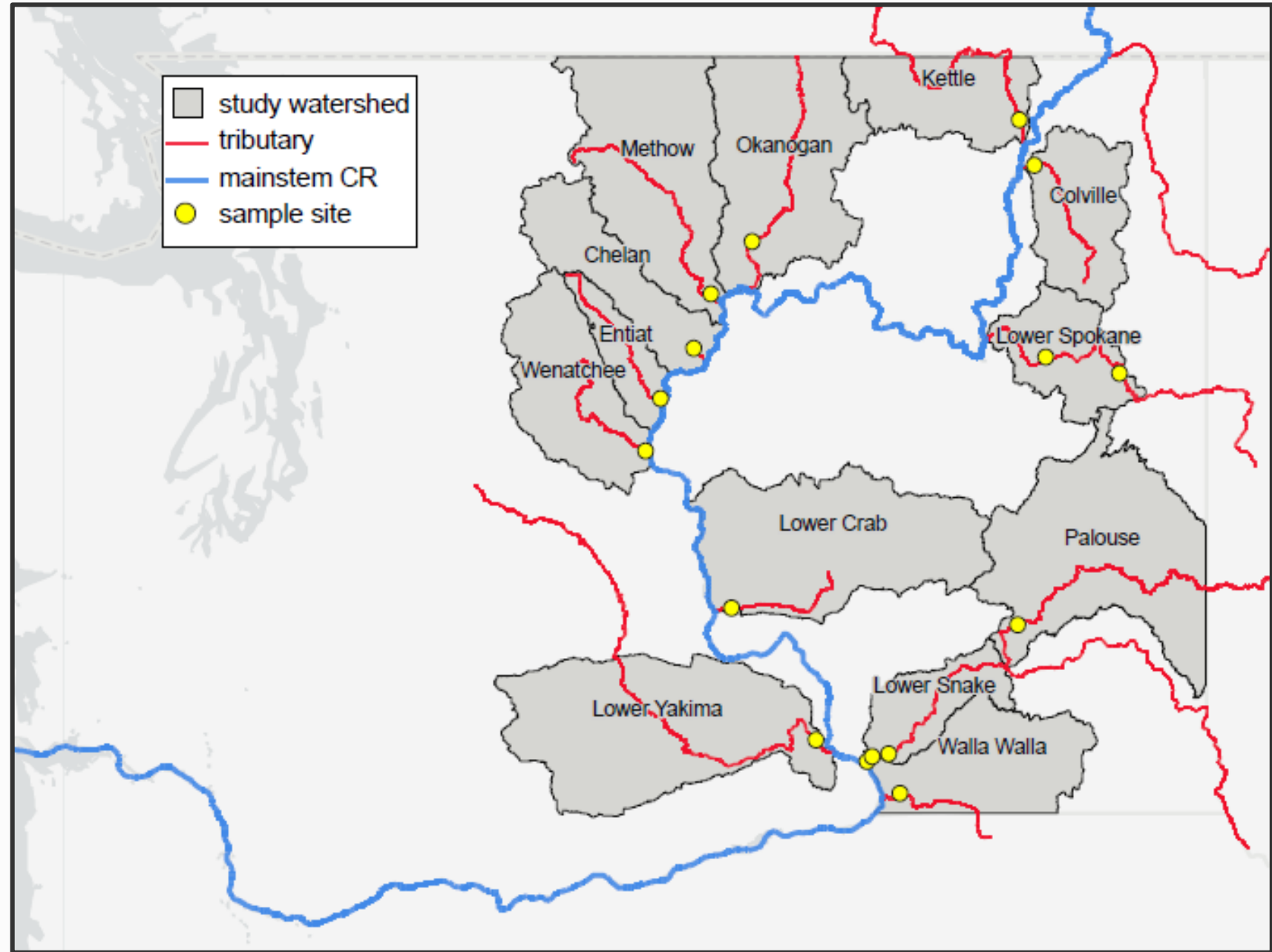
Goal 1

Sample and assess contaminant load and concentrations at a network of monitoring stations in major tributaries to the mainstem Columbia River

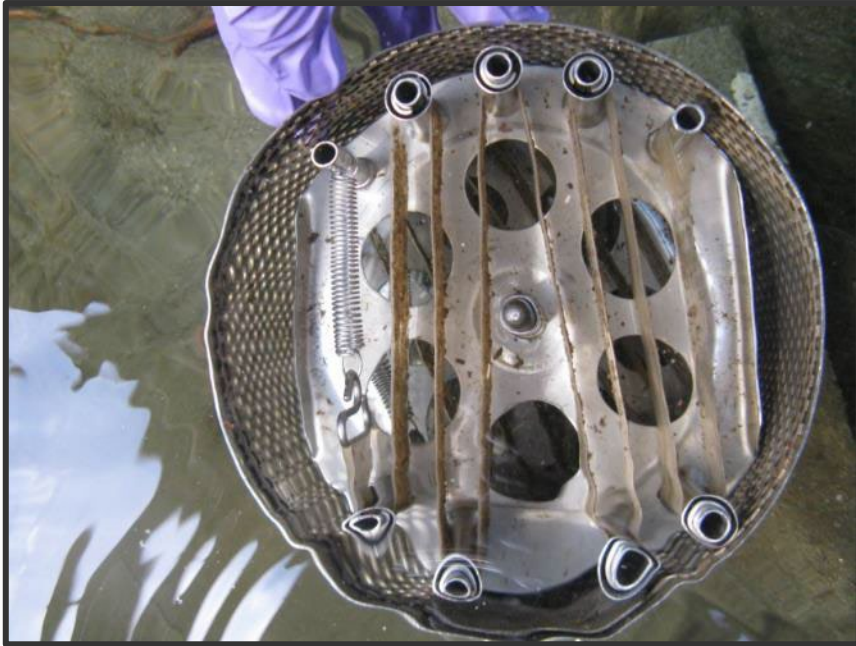
- Major Tributary monitoring stations in the mid and upper-Columbia
- Synoptic survey of inputs (concentrations and loads) to the mainstem at high and low flow – fixed stations with flow gages
- PCBs, PDBEs and chlorinated pesticides (e.g. DDx)
- Establish bioconcentration factors at the lower trophic levels

Sites

- One location per tributary, except Spokane and Snake.
- 2 - 4 sampling events depending on media
- **Passive** – Low Density Polyethylene (water and sediment) and biofilm
- **Active** – large volume samples (200L; particulate and dissolved phase)



Passive Sampling



- Semi-permeable membrane devices (SPMDs)
- Calculated 30-day average water concentration

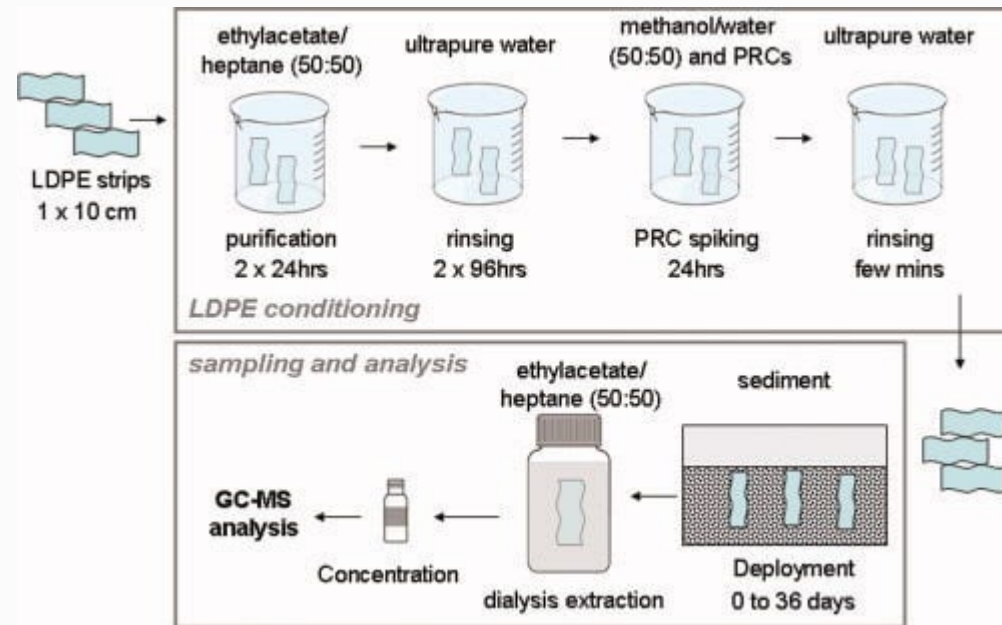
- Biofilm (periphyton, microbial growth and detrital organics)
- Natural passive sampler; base of the river food web



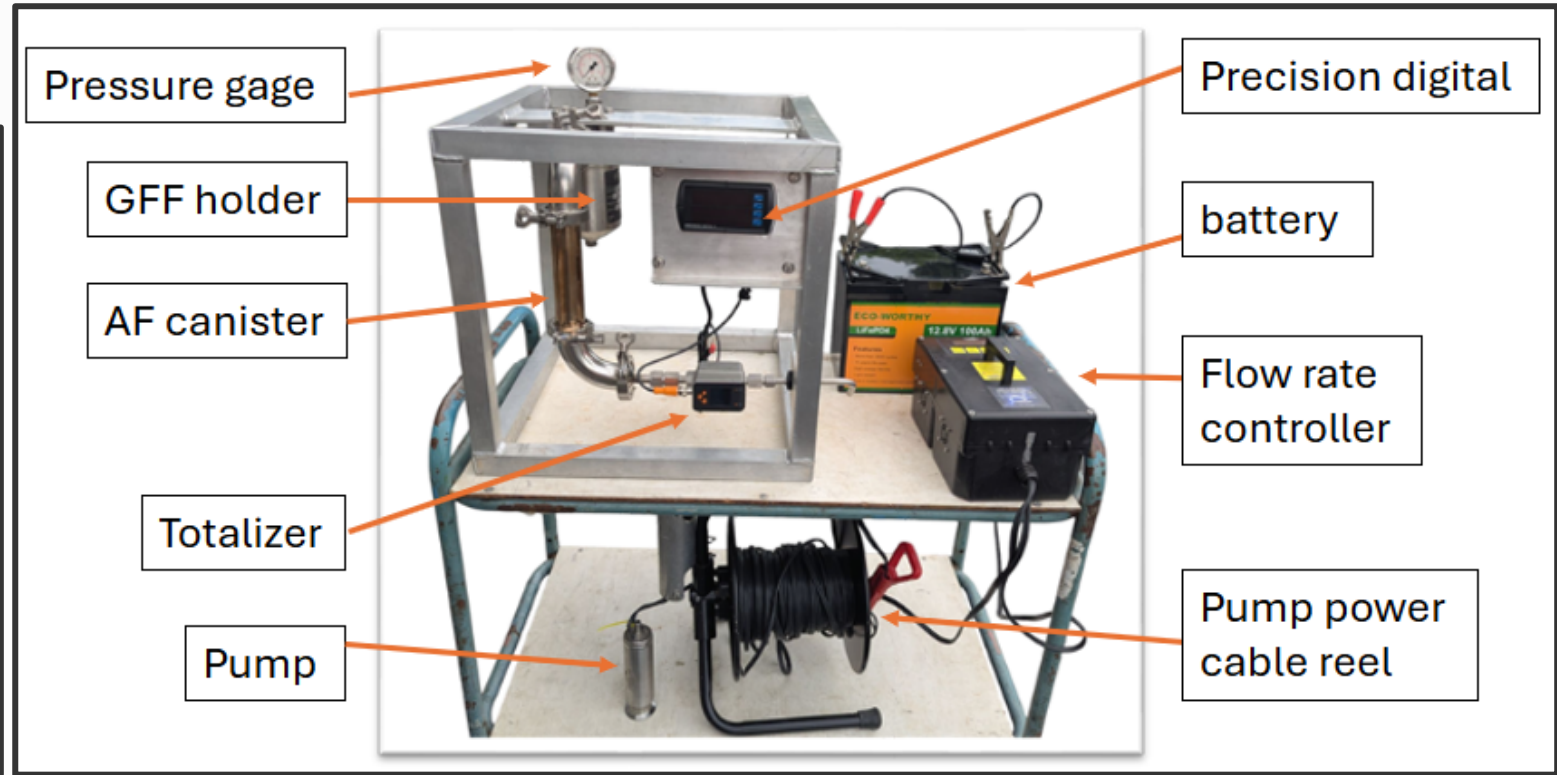
Sediment Sampling



- Upper ~2-3cm composited from 3 independant grab samples
- Water-sediment interface intact
- Split for bulk chemistry and porewater chemistry (ex situ LDPE strips).



Active Sampling



- large volume samples (~200L)
- particulate phase ($>1\ \mu\text{m}$) on GFFs
- colloidal/dissolved phase on PUF media

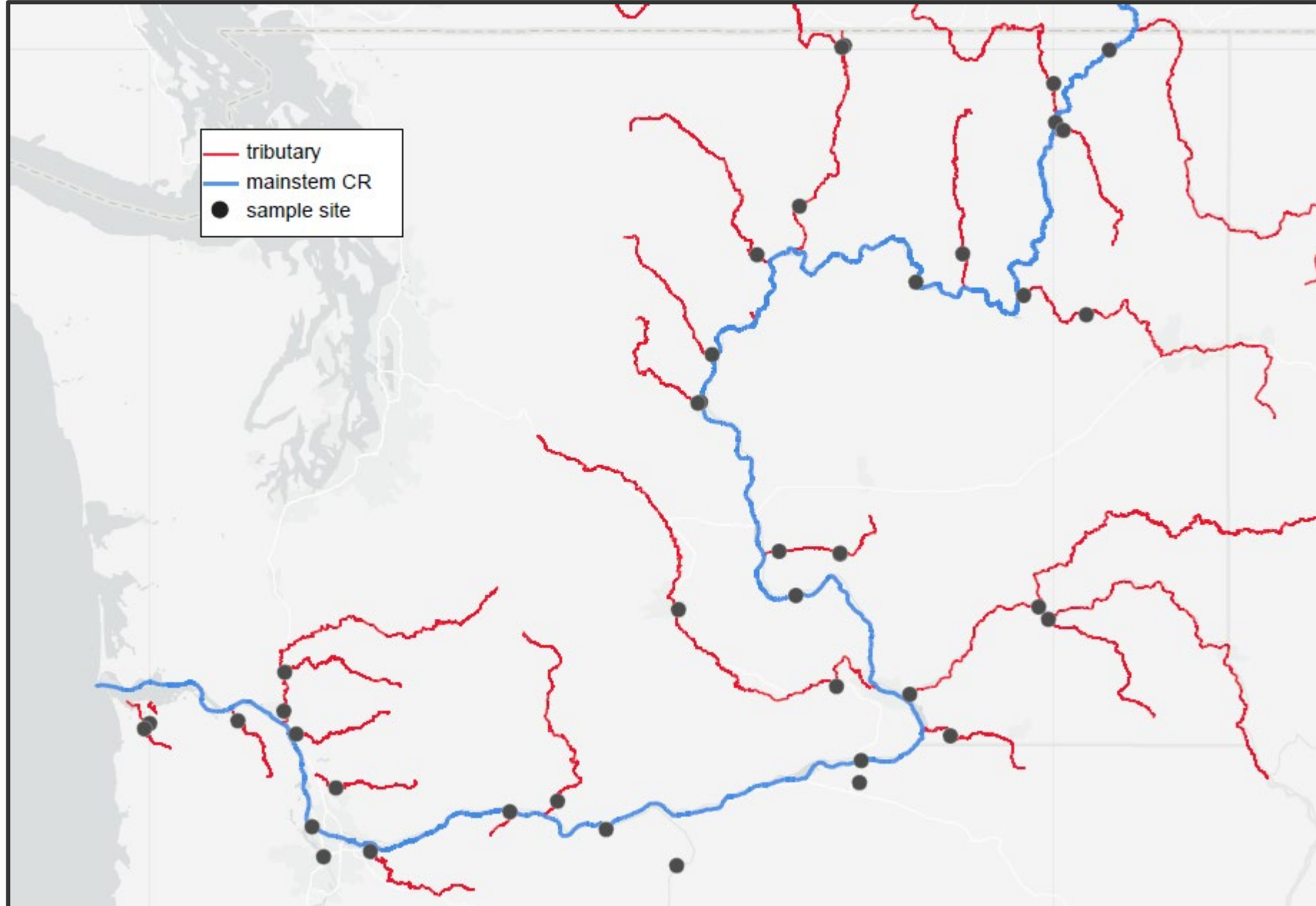
Goal 2

Establish and evaluate a baseline dataset for resident fish otolith microchemistry and fish life history

- Collaboration with WDFW, Otolith and Aging Labs
- Establish the movement and provenance of resident fish tissues used in contaminant monitoring.
- Grab samples at ambient WQ stations to establish a geochemical baseline for otolith microchemistry.
- Use Ecology's Freshwater Fish Contaminant Monitoring Program collections (2024-2027) in the Mainstem Columbia R. to describe the geographic provenance of fish tissues used in toxics monitoring.
- PCBs, PDBEs, chlorinated pesticides (e.g. DDx), Hg and PFAS.
- Pilot study in the Wenatchee Basin with archived fish otoliths

Sites

- Water: 41 sites sampled quarterly for 2 years.
- Triplicate samples for Al, As, B, Ba, Ca, Cu, Fe, K, Li, Mg, Mn, Na, Pb, Rb, Sr, U, V, Zn and Sr isotopes.



Sites

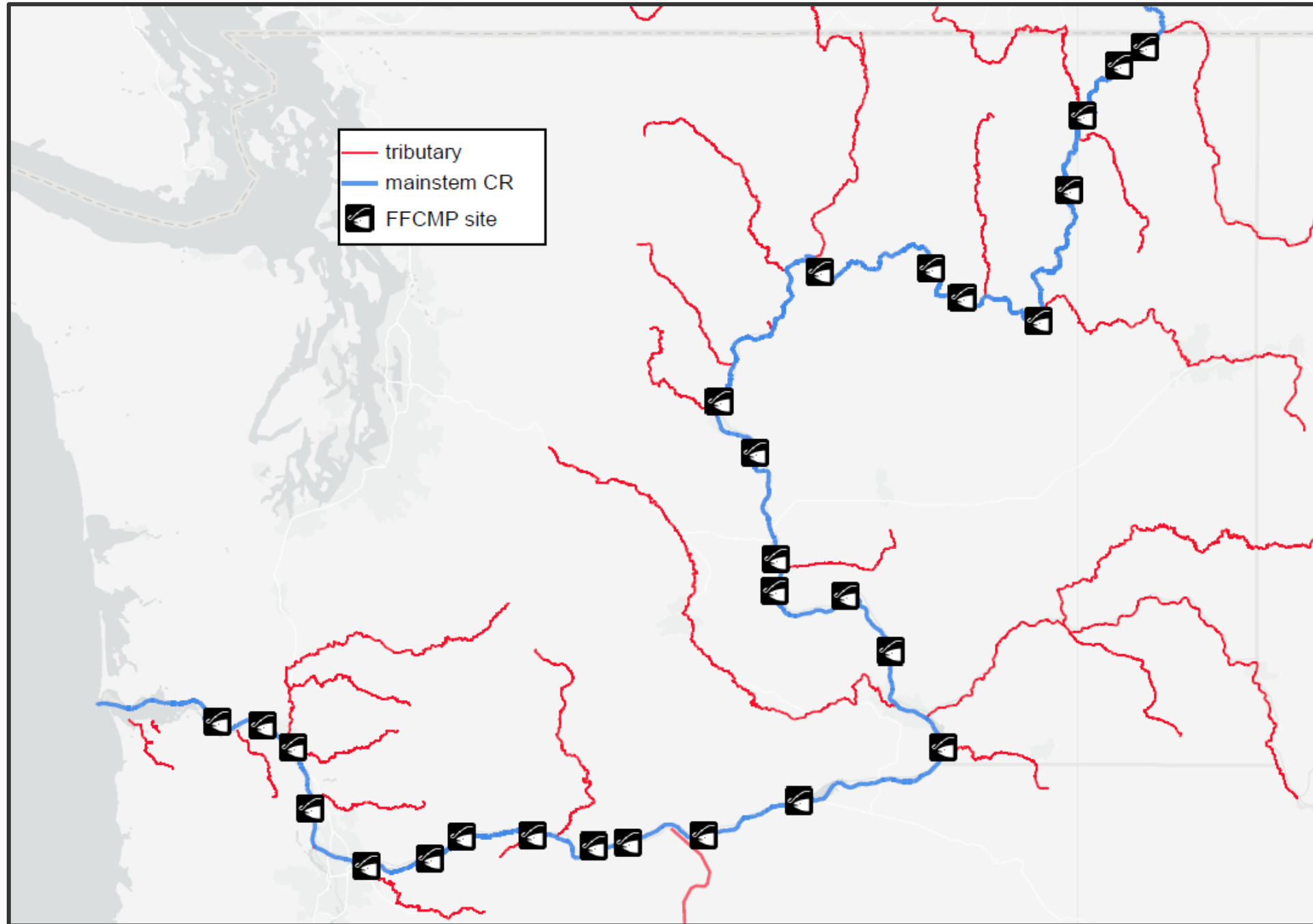
2024: Columbia R-1
(estuary to Bonneville
Dam);

2025: Columbia R-2,
lower (Bonneville
Dam to McNary Dam)

2026: Columbia R-3,
middle (McNary Dam
to Priest Rapids Dam)

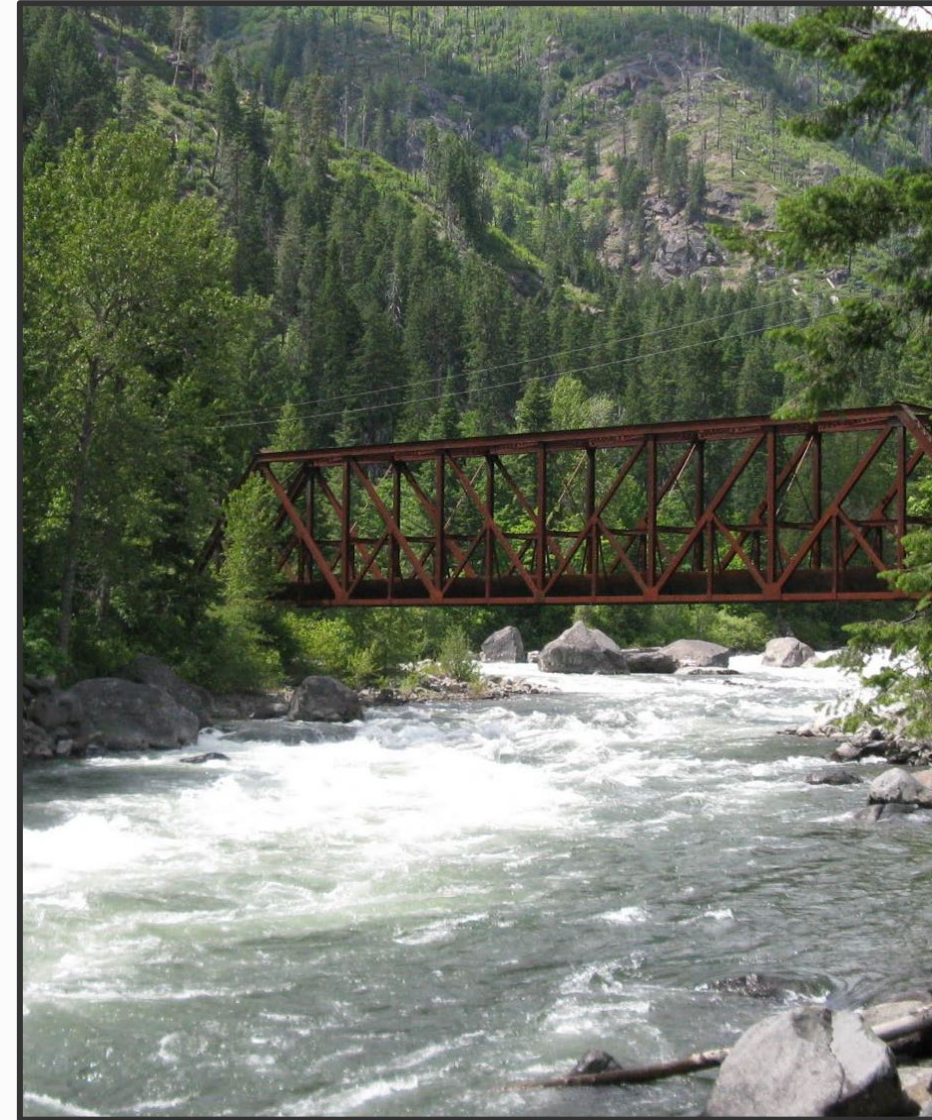
2027: Columbia R-4,
middle (Priest Rapids
Dam to Grand Coulee
Dam)

2028: Columbia R-5,
upper (Grand Coulee
Dam to Northport)



Pilot Study – Wenatchee River

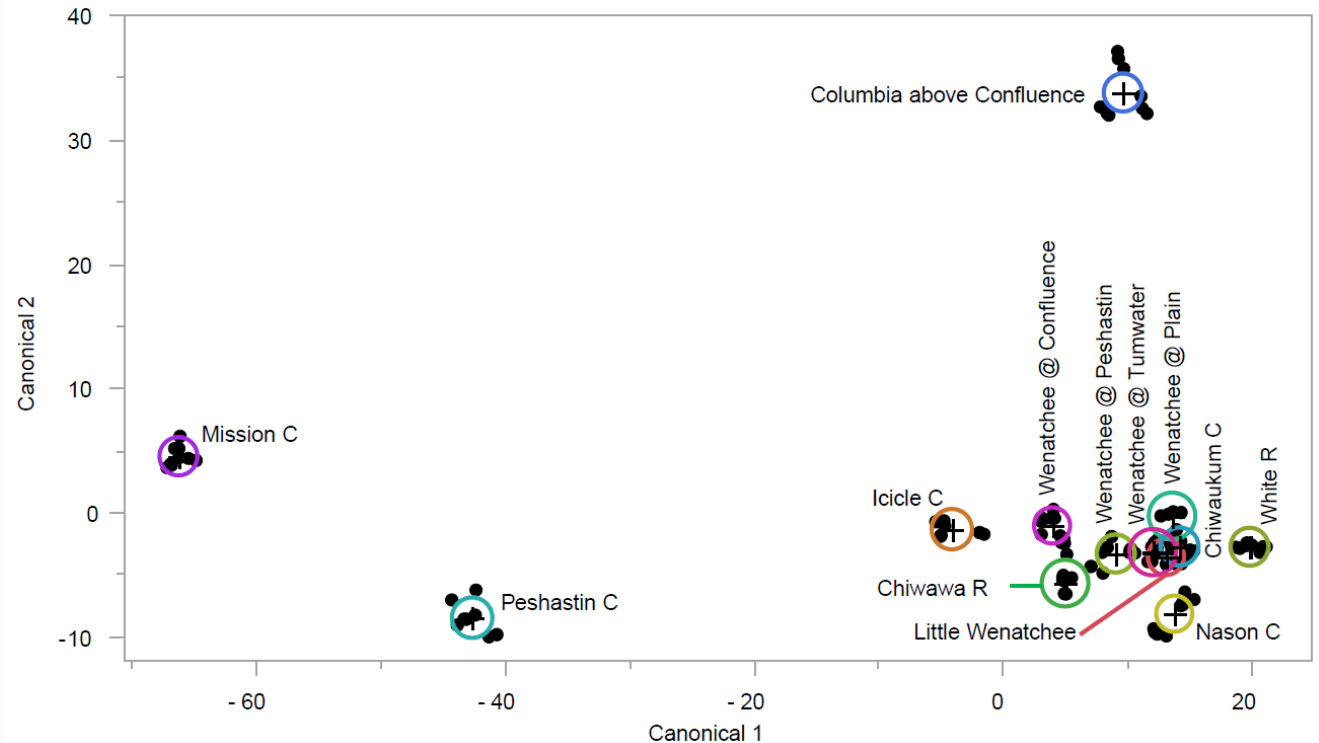
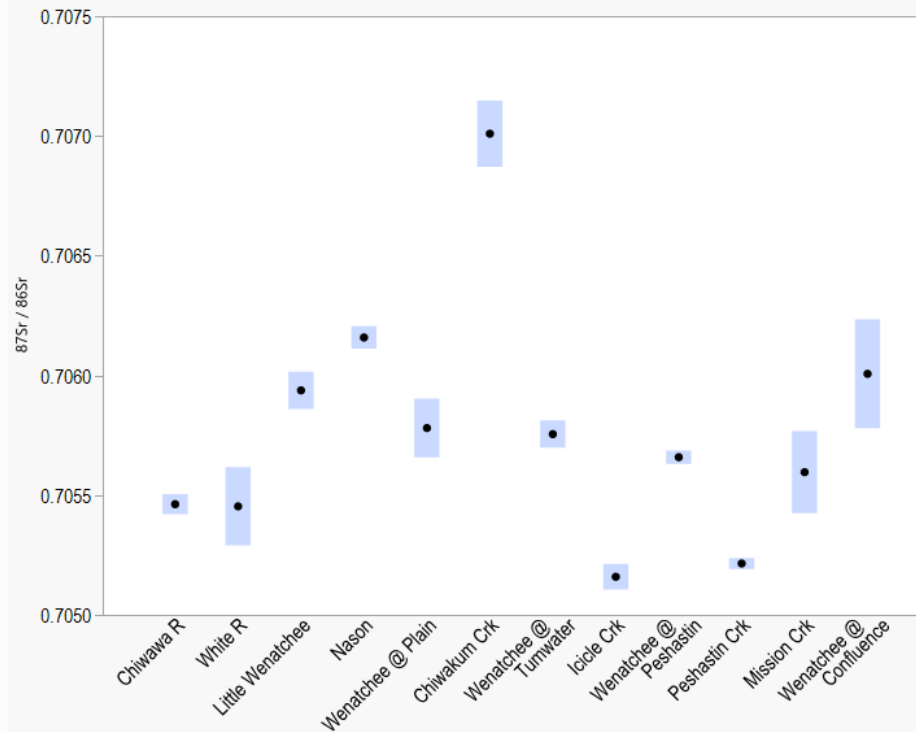
1. Are there clear differences in water chemistry at the HUC10 (tributary) watershed level?
2. Can we thin section resident fish otoliths and get useable microchemistry results?
 - a) Mountain whitefish archived collections (2004, 2010 and 2015)



Water Chemistry

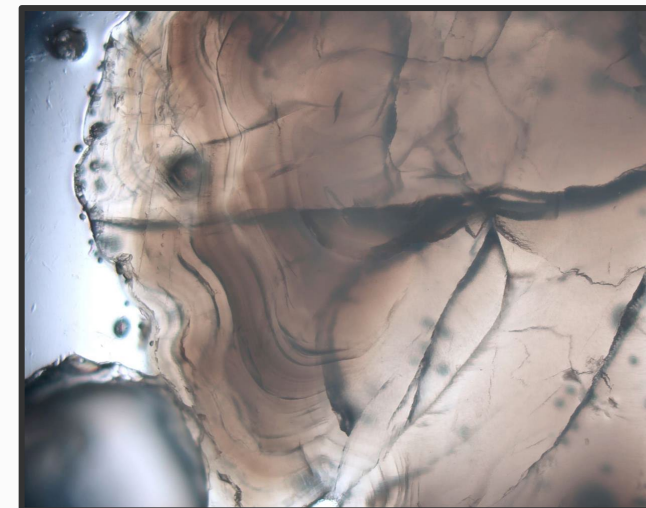
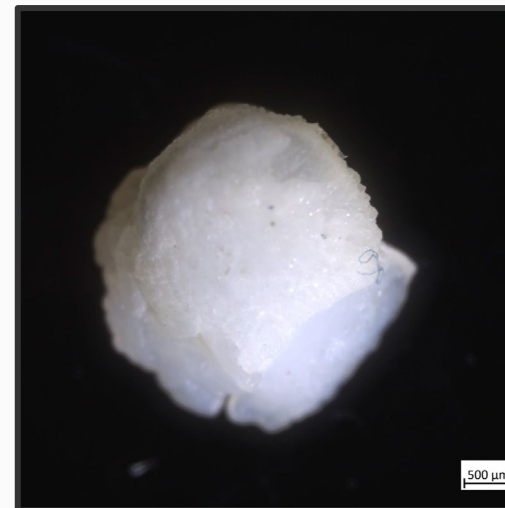
- Verified that samples need to be field filtered; concentrations increase if filtering is delayed.
- Seasonal differences in metals concentrations
 - Necessary to capture various hydrologic regimes
 - Samples will be collected over 2 years during high and low flow periods
- Significant spatial differences between mainstem Wenatchee, Columbia and several tributaries
 - Spatial differences should be resolvable over larger scales throughout CRB

$^{87}\text{Sr}/^{86}\text{Sr} \pm \text{SD}$



Otoliths

- Whitefish otoliths can be thin sectioned and provide adequate annuli for microchemistry
 - Analysis underway at OSU
- Will proceed with analyzing ~100 archived otoliths
- Large scale sucker otoliths (lower Columbia R) may present some complications due to size
 - Typically the operculum is used for ageing
- Bass and walleye otoliths (lower Columbia) are currently being processed



Future Work

- Use this toxics loading study as a prioritization tool
 - Focus on tributary watersheds with high loading of toxics to the mainstem CR for toxics reduction efforts
 - Guide Ecology-led source identification work
- Support the development of a mainstem monitoring program
- Continue to develop and apply tools for understanding resident fish movement



Thank you

William Hobbs

william.hobbs@ecy.wa.gov

Project QAPP: Report 26-03-106

Google: “Ecology Hobbs Columbia”

Project Timeline

Project year	Federal fiscal period	Project planning and manufacturing of equipment	Final published QAPP	Sampling for Toxics	Sampling for metals / isotopes	Project data reduction	Project final report and data archiving
1	10/01/2024 - 03/31/2025						
1	04/01/2025 - 09/30/2025			Aug/Sept 2025			
2	10/01/2025 - 03/31/2026			May/June 2026			
2	04/01/2026 - 09/30/2026			Aug/Sept 2026			
3	10/01/2026 - 03/31/2027			May/June 2027	WY 2027		
3	04/01/2026 - 09/30/2027			Aug/Sept 2027	WY 2027		
4	10/01/2027 - 03/31/2028				WY 2028		
4	04/01/2028 - 09/30/2028				WY2028		
5	10/01/2028 - 03/31/2029						
5	04/01/2029 - 09/30/2029						